PROTOCOL

Cell viability

— Live/dead stain using Invitrogen LIVE/DEAD Cell Imaging Kit

Application

This protocol explains how to perform Cell viability using Invitrogen LIVE/DEAD Cell Imaging Kit with P3D Scaffolds. For more protocols, please visit our Resources Platform.

Materials

- Invitrogen LIVE/DEAD cell imaging kit
- Fresh media
- Microscope with FTIC/GFP and Texas Red filter
- Tinfoil for covering the samples during incubation time

Flowchart

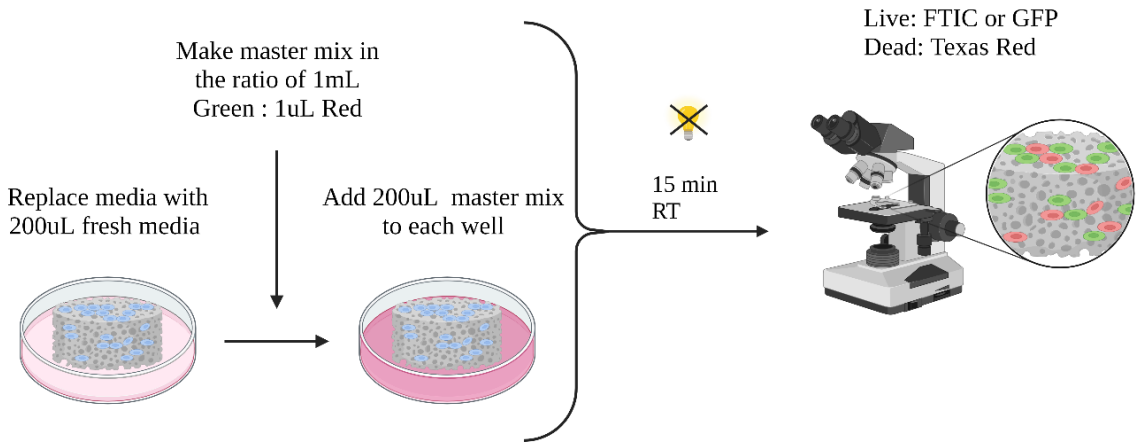

FIGURE 1: *WORKFLOW FOR EVALUATING CELL VIABILITY USING THE LIVE/DEAD CELL IMAGING KIT BY INVITROGEN.*

Notes before starting and general advice on material handling

- All handling of the P3D Scaffolds should be performed using gloves, according to the standard aseptic methods.
- The scaffolds are supplied sterile by dry heat sterilization and remain sterile until opened.

Procedure

1. Thaw the cell kit at room temperature.
2. Aspirate the media from the scaffold.
3. Add 200 µL fresh media to each scaffold.
4. Mix the Live/Dead stain master mix by adding 1 mL green staining solution to the 1 µL red staining solution – enough for 5 Ø12mm P3D Scaffold samples. Please upscale the master mix according to your experimental setup.
5. Add 200 µL from the Live/Death cell kit solution to each scaffold.
6. Cover with tinfoil paper to prevent it from light damage.
7. Incubate for 15 min at room temperature.
8. Perform the analyses using a fluorescent microscope.
    a. For the live cells, use FTIC or GFP filters.
    b. For the dead cells, use the Texas Red filters.

Disclaimer:
The products are “For Research Use Only (RUO)” and should not be used for clinical purposes. Ossiform makes no other warranties, expressed or implied, including the implied warranty of merchantability and the implied warranty of fitness for particular purpose.

Patent status:
The product is protected by one or more US, European, and/or foreign patents.

Safety Data Sheet:
Download from our website here.

Technical Data Sheet:
Download from our website here.