

## PROTOCOL

### Cell viability

– Live/dead stain using Invitrogen LIVE/DEAD Cell Imaging Kit

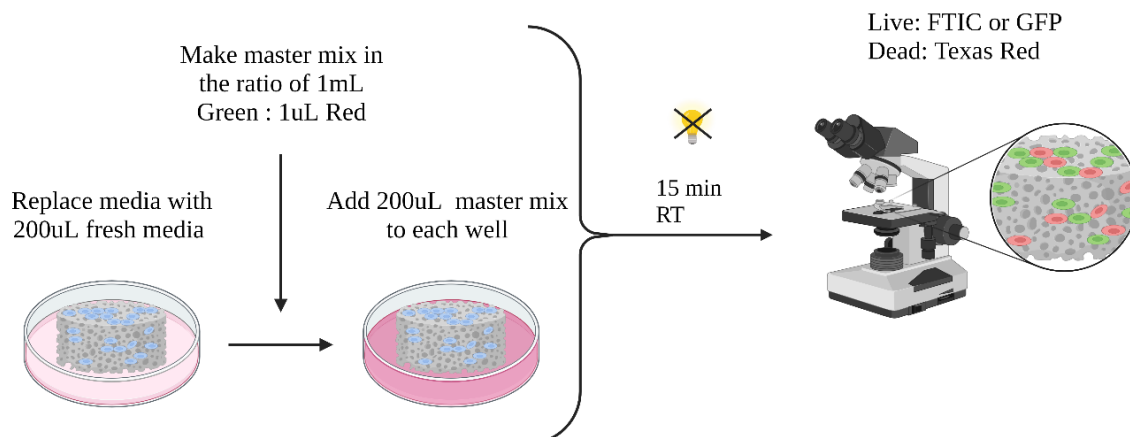
#### Application

This protocol explains how to estimate cell viability using Invitrogen LIVE/DEAD Cell Imaging Kit with P3D Scaffolds. For more protocols, please visit our [Resources Platform](#).

#### Materials

- Invitrogen LIVE/DEAD cell imaging kit
- Fresh media
- Microscope with FTIC/GFP and Texas Red filter
- Tinfoil for covering the samples during incubation time

#### Flowchart



**FIGURE 1:** WORKFLOW FOR EVALUATING CELL VIABILITY USING THE LIVE/DEAD CELL IMAGING KIT BY INVITROGEN.

## Notes before starting and general advice on material handling

- All handling of the P3D Scaffolds should be performed using gloves, according to the standard aseptic methods.
- The scaffolds are supplied sterile by dry heat sterilization and remain sterile until opened.

## Procedure

1. Thaw the cell kit at RT.
2. Aspirate the media from the scaffold.
3. Add 200 µL fresh media to each scaffold.
4. Mix the Live/Dead stain master mix by adding 1 mL green staining solution to the 1 µL red staining solution – this should be enough for 5 Ø12mm P3D Scaffold samples. Please upscale the master mix according to your experimental setup.
5. Add 200 µL from the Live/Death cell kit solution to each scaffold.
6. Cover with tinfoil paper to prevent it from light damage.
7. Incubate for 15 min at RT.
8. Perform the analyses using a fluorescent microscope.
  - a. For the live cells, use FTIC or GFP filters.
  - b. For the dead cells, use the Texas Red filters.

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**Technical Data Sheet:**

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